

# Diversity above and below ground: the genetics of vernal pool plants and seed banks

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# The problem:

- Understanding vernal pool (or any) plant communities is important...
- ...but hard!
  - time consuming
  - expertise
  - temporal variance
  - issues with ID'ing degraded/dried out samples
  - Can't ID below-ground species





# Use eDNA and metabarcoding genomic techniques to:

- ▶ ID all plant species present
  - ▶ detect T&E species
- ▶ characterize plant spp genetically
- ▶ Reduce site visits
- ▶ Increase accuracy
- ▶ Increase understanding of VP communities
- ▶ Above/below ground ("seed bank") diversity
- ▶ Apply method to other soil systems



*Neostapfia colusana*  
"Colusa grass"  
Fed: Endangered  
CA: Threatened

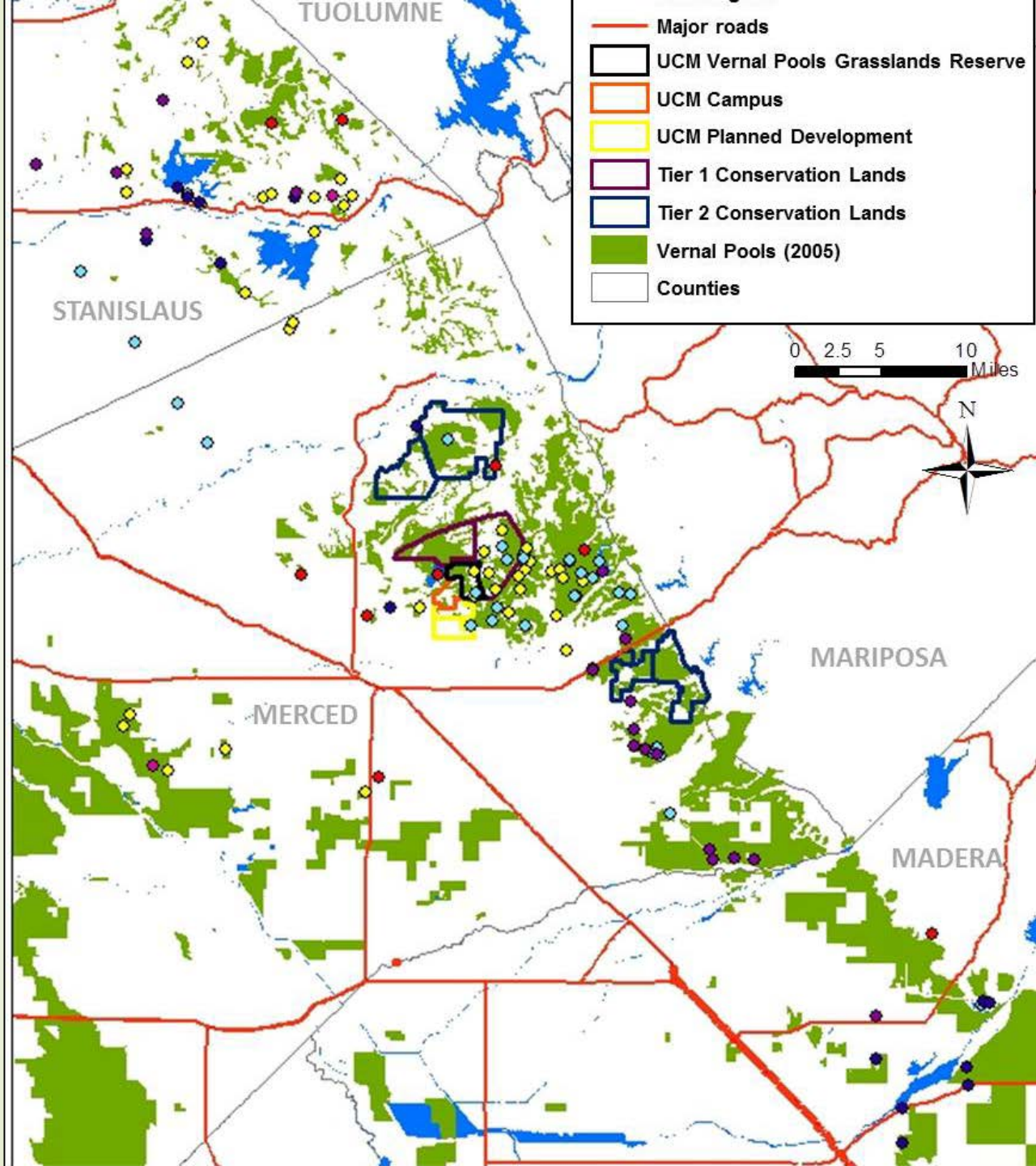


*Orcuttia inaequalis*  
"San Joaquin Valley Orcutt Grass"  
Fed 'Threatened  
CA Endangered



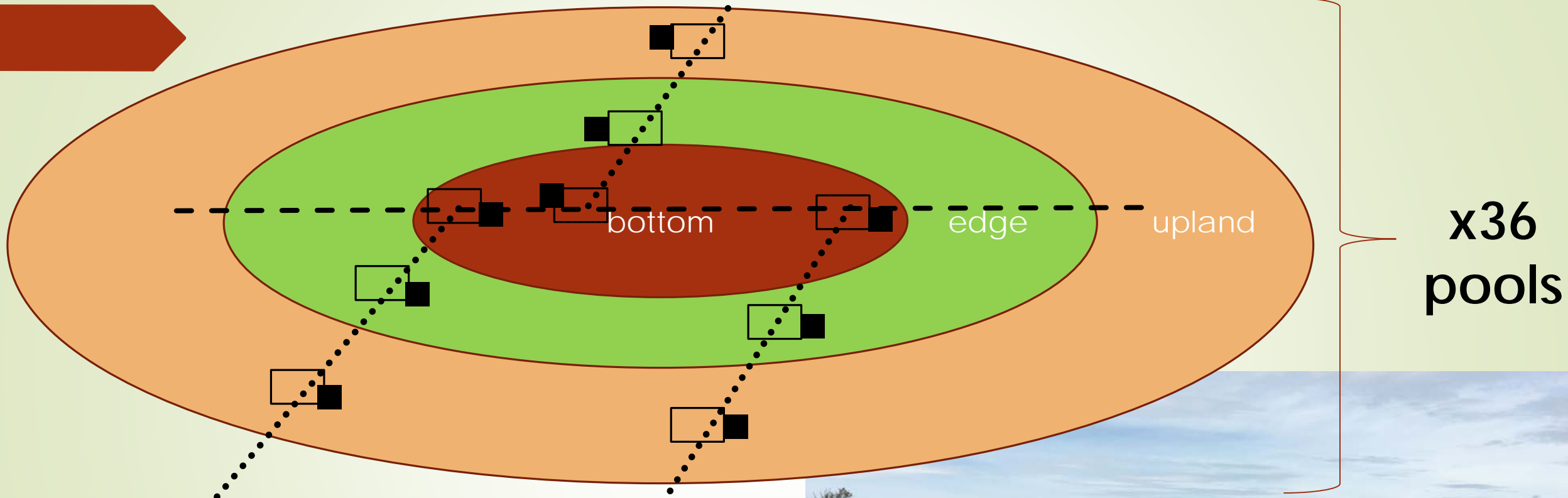
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*Orcuttia pilosa*  
"Hairy Orcutt Grass"  
Fed Endangered  
CA Endangered





# MVPGR veg. surveys and "seed bank" sampling

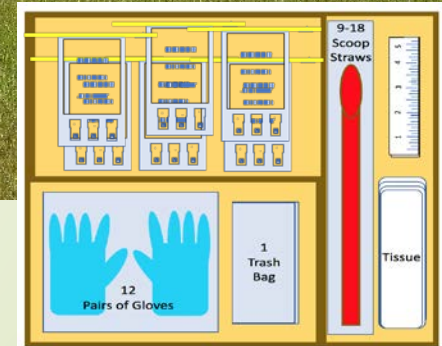


- Quadrat-transect surveys (above ground, % cover)
- Soil sampling (below ground, 4x4x2")



# California environmental DNA

- ▶ establish a baseline of California's biodiversity
  - ▶ environmental DNA museum
  - ▶ Temporal replication
- ▶ Citizen science



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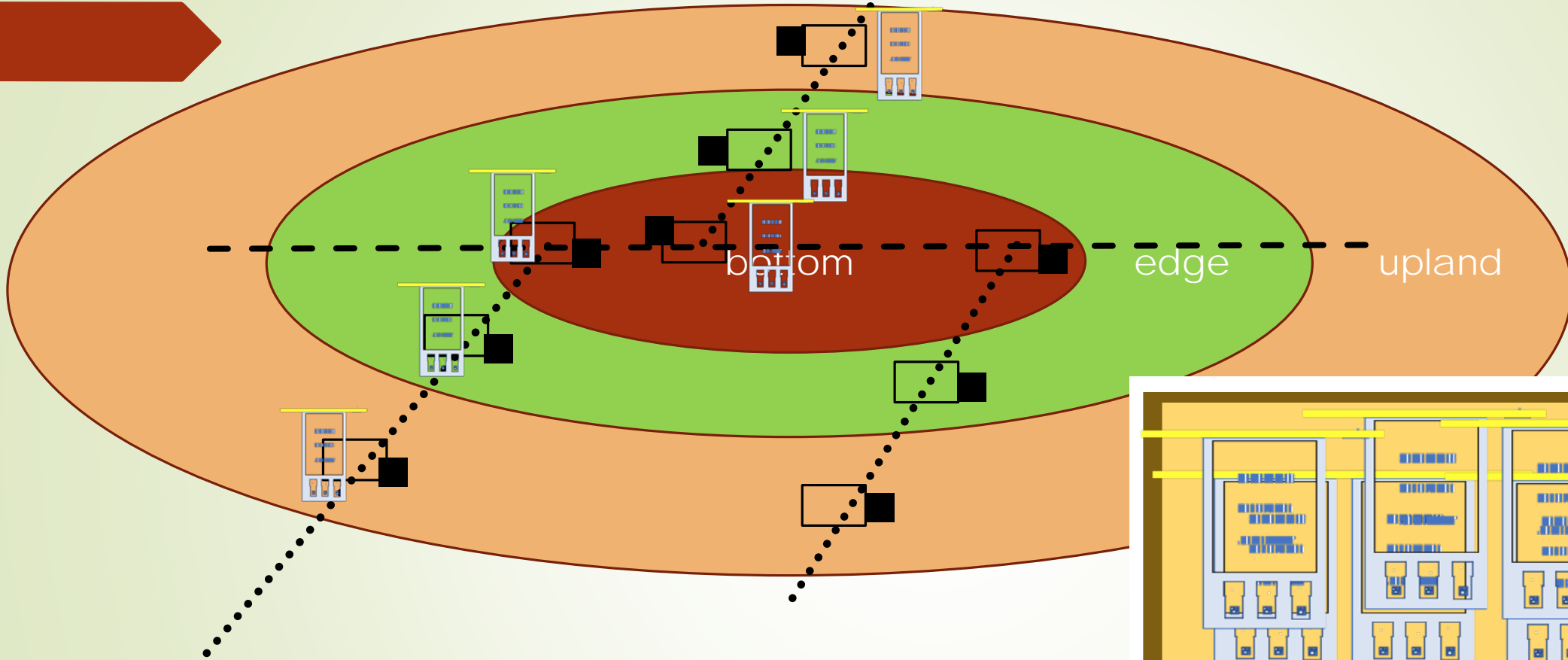


CALeDNA

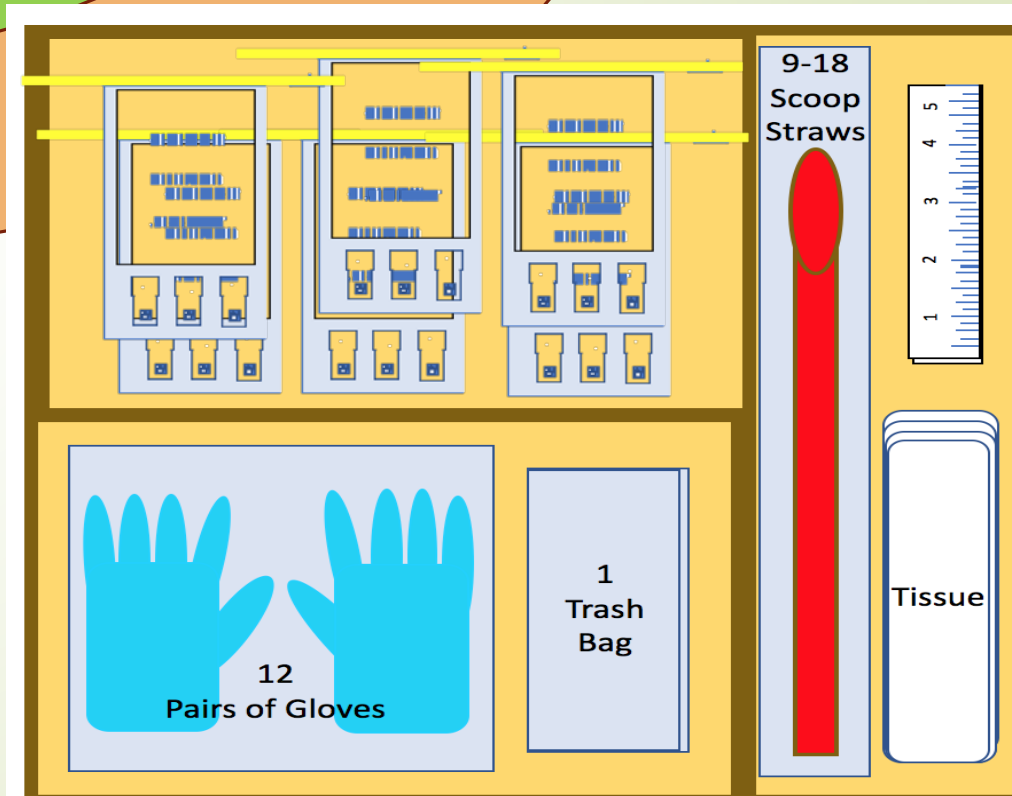
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# CaleDNA soil sampling

(5 pools)



=3 tubes, each zone; 18 tubes total per pool





# Pilot: Compare Soil Sample Processing

- |                                       |               |           |
|---------------------------------------|---------------|-----------|
| 1. Germinate in greenhouse            | → ID species  | } Compare |
| 2. Sieve soil, extract, metabarcode   | → ID species* |           |
| 3. Grind soil, extract, metabarcode   | → ID species* |           |
| 4. Pool samples, extract, metabarcode | → ID species* |           |
| 5. CaleDNA samples                    | → ID species* |           |

- \*Metabarcoding sequencing (Illumina miSeq)
  - COI, ITS2, rbcL and trnL
  - CaleDNA -- ITS1 universal primer and CO1 primer
- "known" seed bank samples





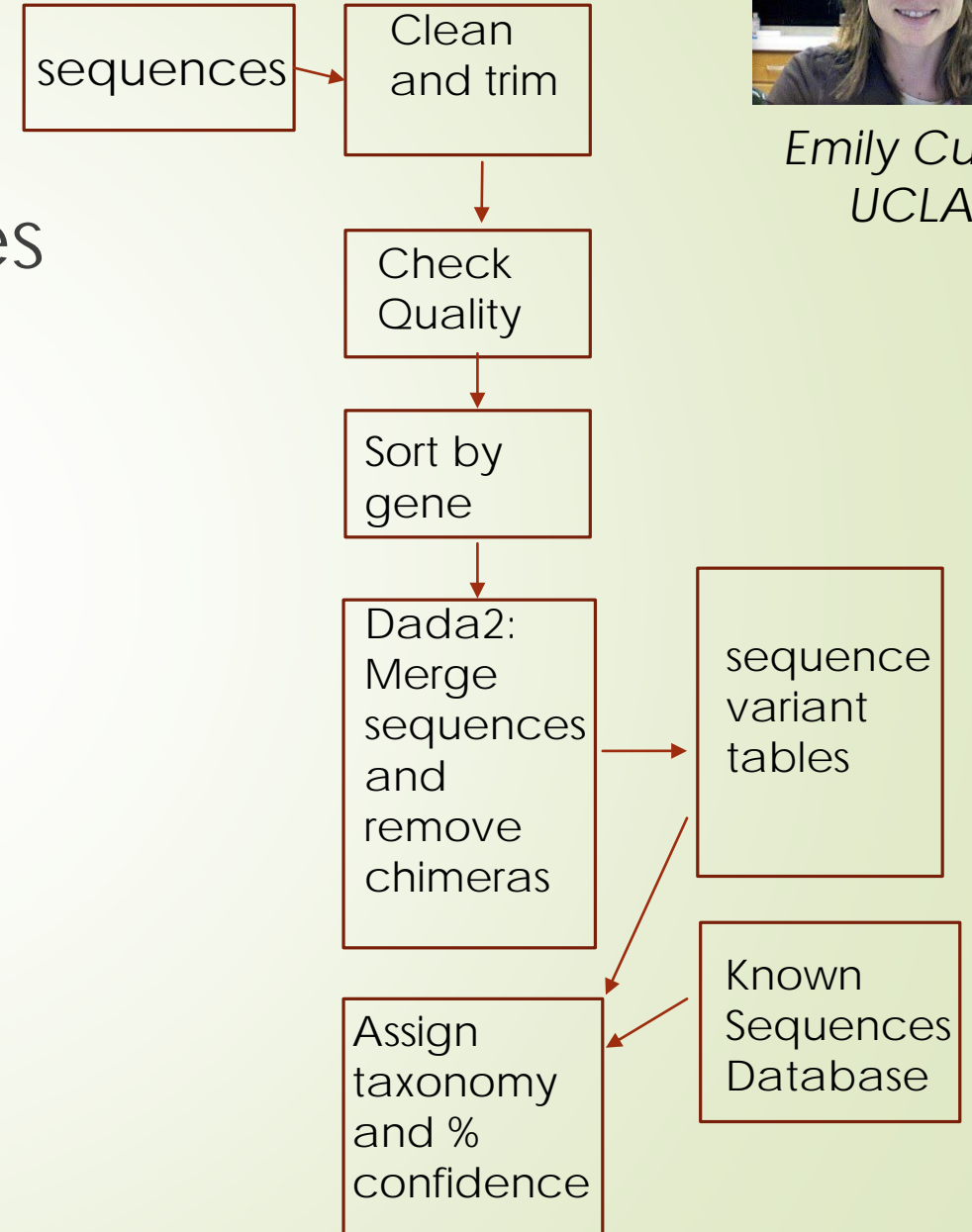
# eDNA for VP soils



- Species ID (T&E) and detection rates (at known T&E plant sites)
- $\beta$  (species) diversity
- $\pi$  (genetic) diversity; pop. structure
- Environment:
  - hydrology (inundation, pool connectivity)
  - soil type
  - Climate
  - Spatial structure

# Methods

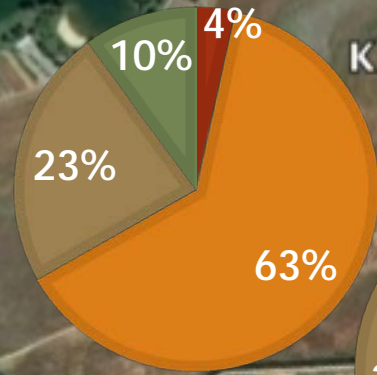
- DNA extracted from soil samples
- 5 genes:
  - 16S - prokaryotes
  - CO1- eukaryotes
  - 18S - eukaryotes
  - ITS1 - plants
  - ITS2 - fungi
- ANACAPA bioinformatics pipeline



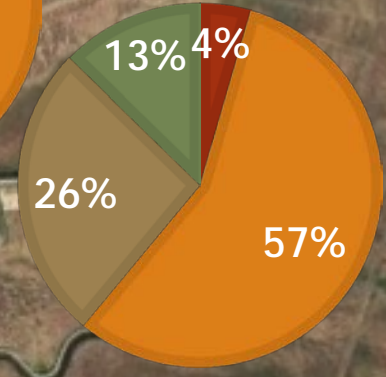
Emily Curd,  
UCLA



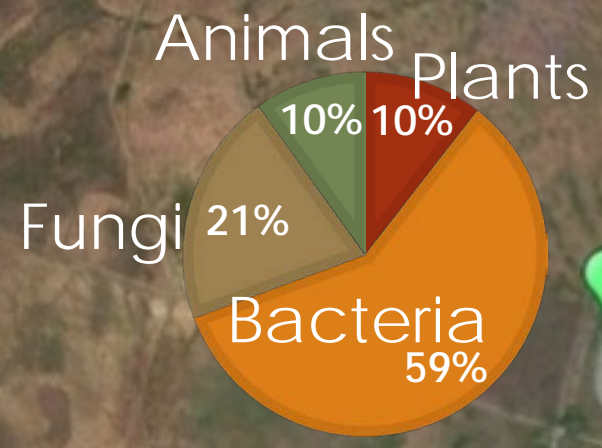
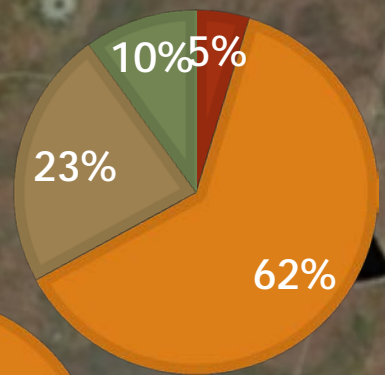
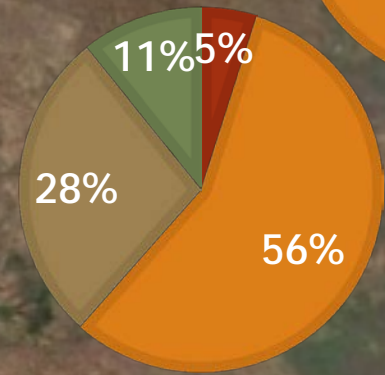
# Proportion of species found



K110  
K112  
K124

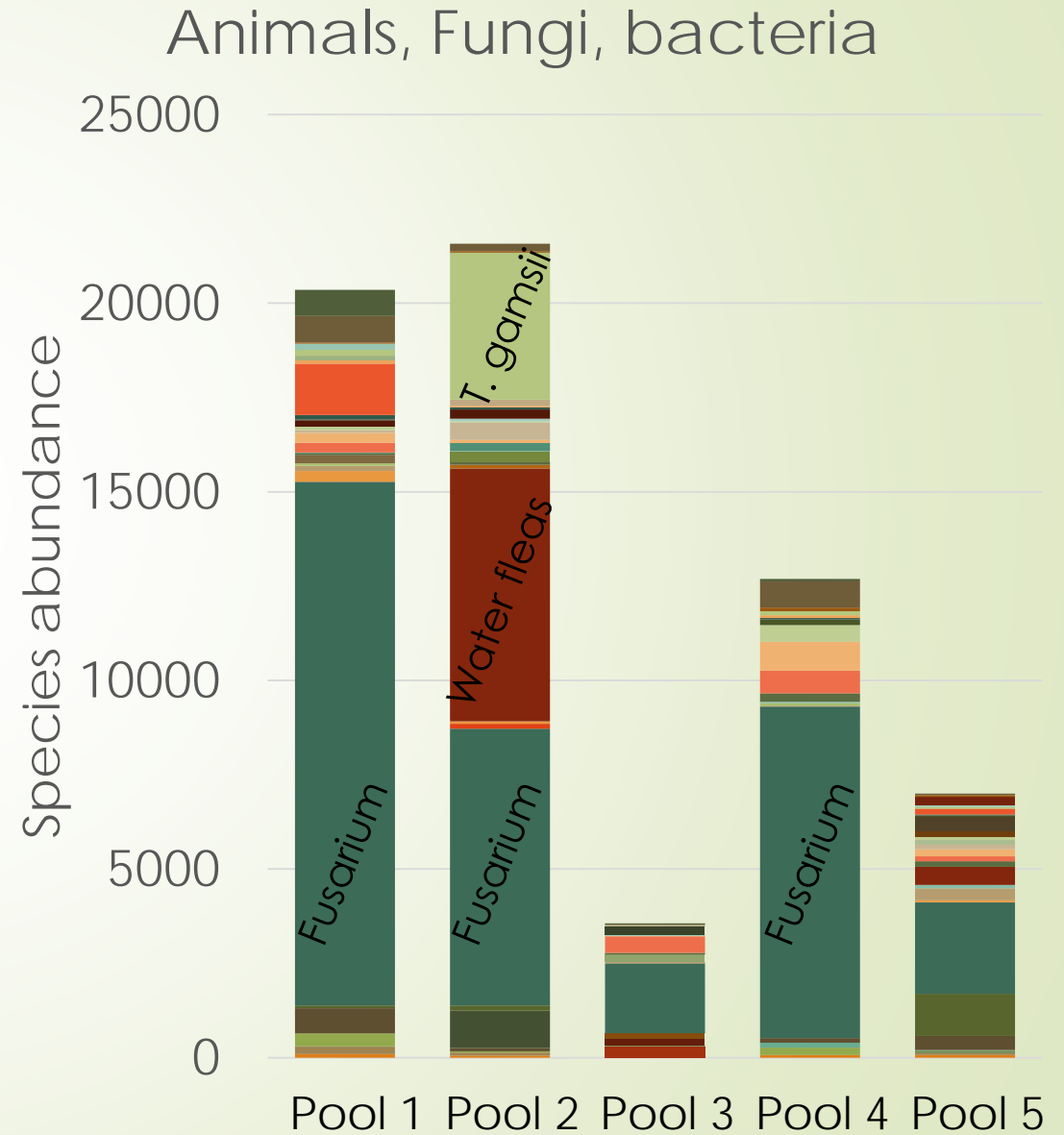
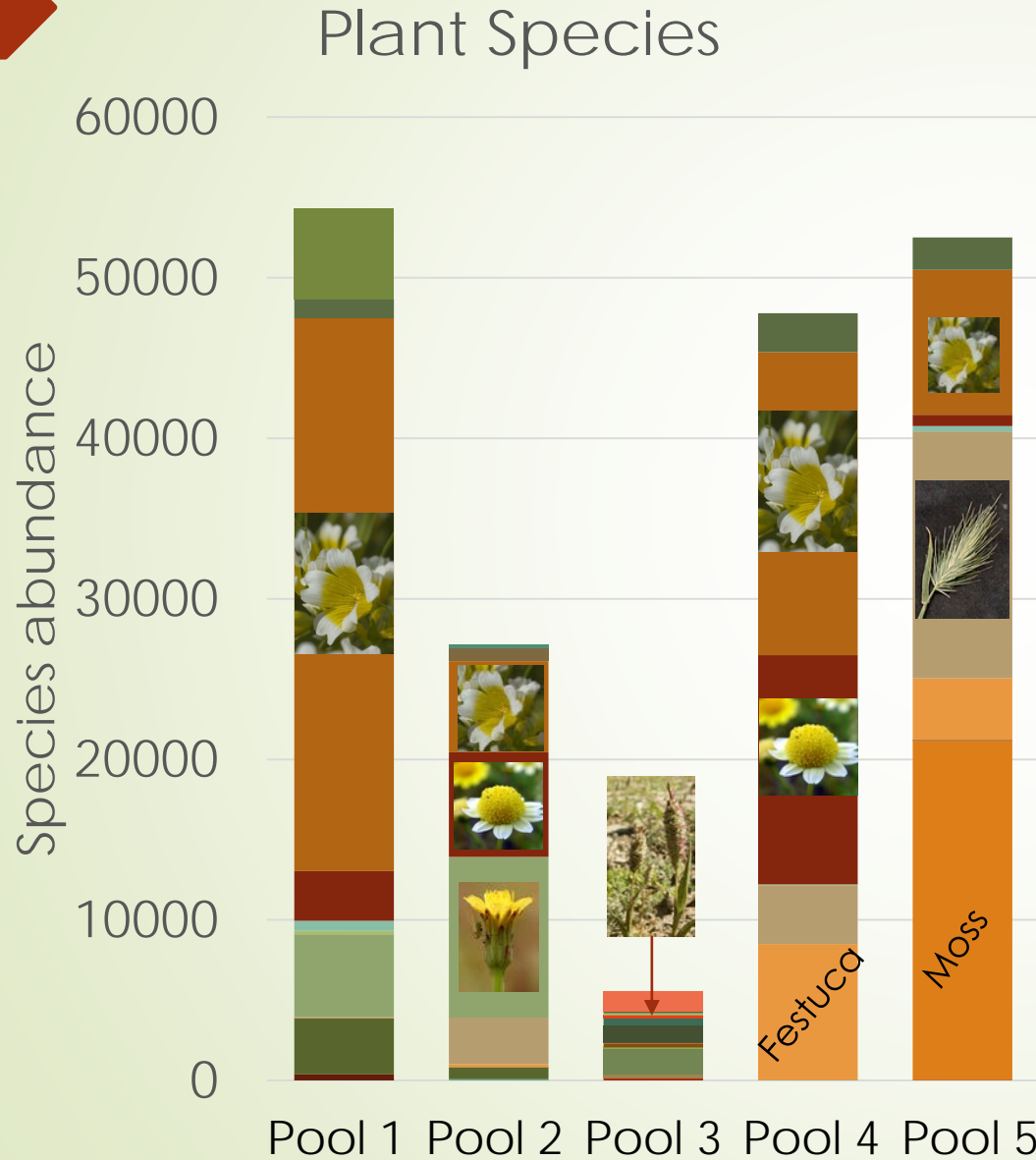


K112  
K129



South Playa

# Species abundance and composition



^ South Playa Pool, *N. colusana*



# Plant species detected in soil samples using eDNA

## ➤ Native and common vernal pool species



➤ *Lasthenia fremontii* - Frémont's goldfields

➤ *Limnanthus douglasii* - meadowfoam

➤ *Elatine californica* - CA waterwort

## ➤ Endangered and rare species



➤ *Neostafia colusana*

## ➤ Invasive non-native species

➤ *Hordeum marinum*, *Festuca bromoides*

# Community structure – pool connectivity

Bacteria

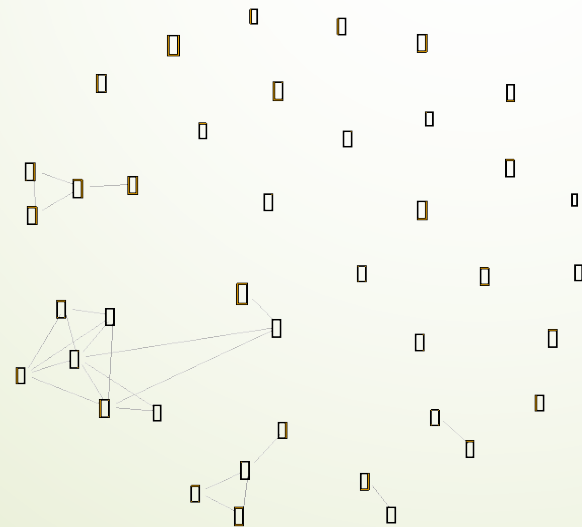


Fungi

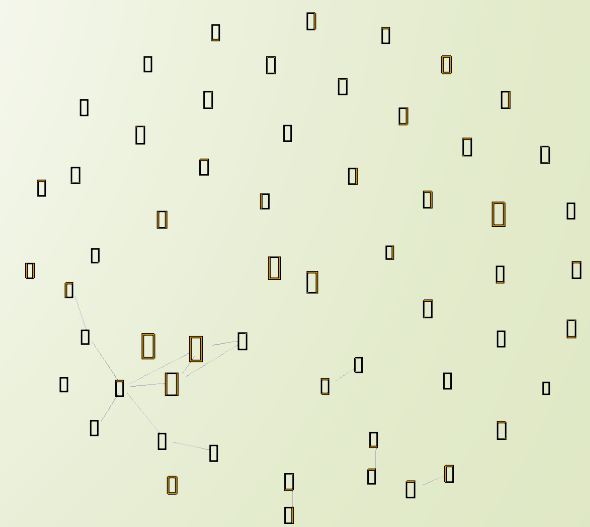


● Species  
— Interaction

Plants and algae



Animals





# Preliminary results: species diversity indices, 5 CaleDNA pool soils

$\beta$  = the ratio between regional and local species diversity  $\beta = \gamma/\alpha$   
 $\alpha$  = mean species diversity in sites or habitats at a local scale  
 $\gamma$  = total species diversity in a landscape  $\gamma = \alpha * \beta$

- Greater spp diversity in bottom, decreasing toward uplands
- Similar patterns for two different markers and classes of organisms

	Plant ITS			COI		
	bottom	edge	upland	bottom	edge	upland
$\beta$	2.5	2.2	1.8	2.3	2.5	2.2
$\alpha$	13.8	12.6	11.6	20.9	18.2	14.5
$\gamma$	48.0	41.0	33.0	70.0	64.0	47.0

# Limitations of eDNA for species detection

- Detection constrained by reference sequence database
  - *Plagiobothrys linifolius*
  - *Poa xenica*
- Sampling effort and area of the pool
- Duration of DNA in the environment
  - *N. colusana* detected before the growing season

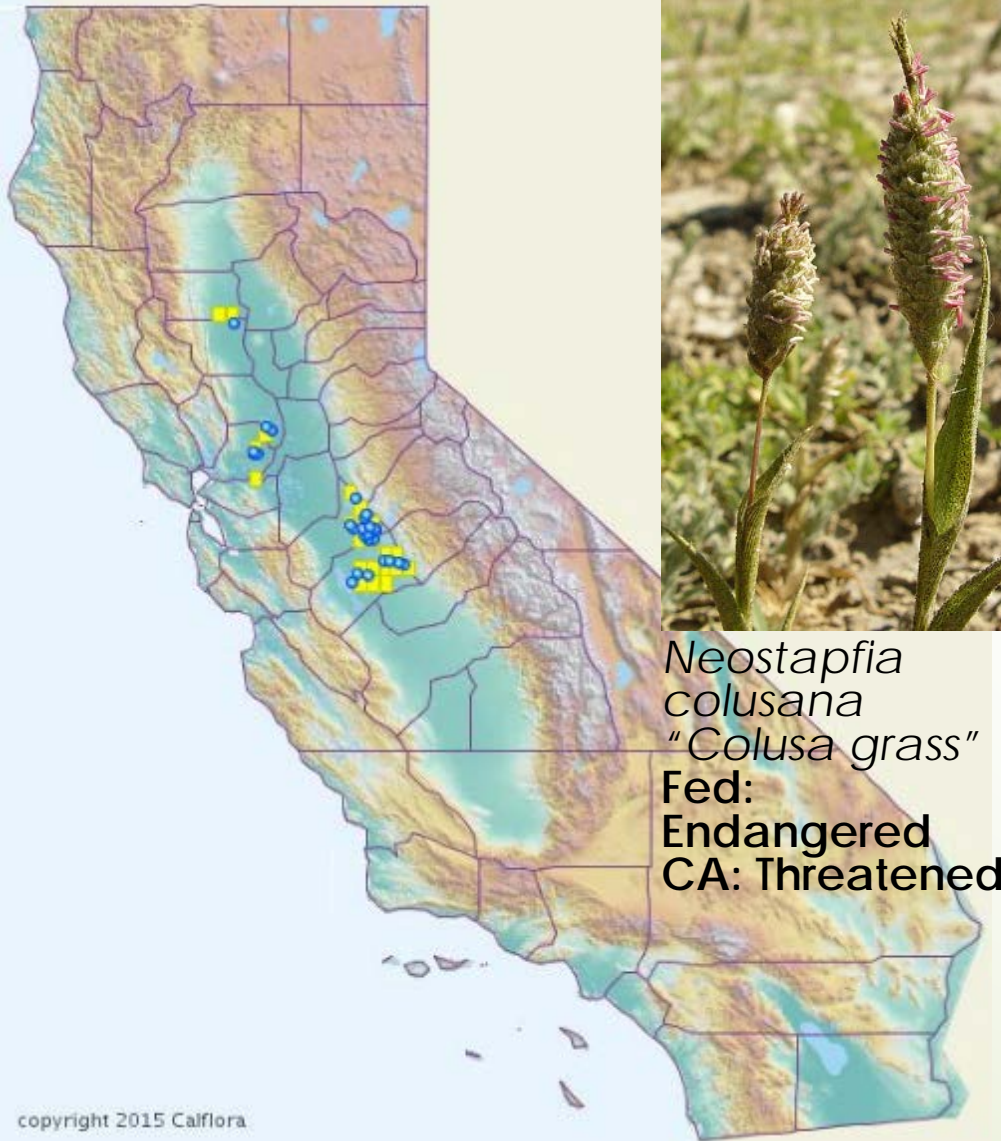




## Next steps

- Improving reference sequence database for vernal pool plants
- Analyze biodiversity above vs below ground
- Community assembly

# Statewide *Orcuttiae* SNP marker development: *Neostapfia colusana* population genetic structure



*Neostapfia colusana*  
"Colusa grass"  
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*Orcuttia inaequalis*  
"San Joaquin Valley Orcutt Grass"  
Fed 'Threatened  
CA Endangered



*Orcuttia pilosa*  
"Hairy Orcutt Grass"  
Fed Endangered  
CA Endangered



*Tuctoria greenei*  
"Greene's tuctoria"  
Fed Endangered  
CA Rare





# Statewide *Orcuttia* SNP marker development: *Neostapfia colusana* population genetic structure

## ➤ RADseq

- SNP marker development for four *Orcuttia* species
- population genetic investigation of *N. colusana* rangewide population structure and diversity
- How many “populations” exist in these isolated pools, how is diversity structured, and how much genetic diversity is left in this threatened species

## ➤ Genome Sequencing

- sequenced *N. colusana* genome (UC Genomics Consortium)
- 2.2GB
- 87% completeness (core genes)
- Next steps: Annotation

# Acknowledgments

## ► Funding

- USBR/USFWS CVPCP grant: "Genetic investigation of listed vernal pool plants and their communities in Merced County"
- UC Regents Catalyst grant; CaleDNA program and citizen scientists: "Conservation genomics of California vernal pool endemics"

## ► Assistance

- Sexton Lab, undergraduate assistants
- Dawson Lab
- UC Conservation Genomics Consortium
  - Emily Curd, Rachel Meyer
  - Citizen scientists
- Carol Witham, Bob Holland, David Muths
- Monique Kolster, MVPGR



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## BACKGROUND

- ▶ USBR/USFWS CVPCP “Genetic investigation of listed vernal pool plants and their communities in Merced County”
  - ▶ improved protocols for soil plant species ID from soil samples
  - ▶ species diversity/richness estimates and environmental correlates
  - ▶ SNP marker development for Orcuttieae (four species)
  - ▶ population genetic investigation of a threatened vernal pool species (*Neostapfia colusana*)
  - ▶ Management implications
- ▶ UC Catalyst program – UC Conservation Genetics Consortium  
CaleDNA “Conservation genomics of California vernal pool endemics”
  - ▶ Genome sequencing for *N. colusana*
  - ▶ Soil eDNA sample collection and metabarcode species ID (plants + other)